REVIEW ARTICLE

Edward W. Campion, M.D., Editor

Pelvic Inflammatory Disease

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ELVIC INFLAMMATORY DISEASE IS AN INFECTION-INDUCED INFLAMMATION of the female upper reproductive tract (the endometrium, fallopian tubes, ovaries, or pelvic peritoneum); it has a wide range of clinical manifestations.¹ Inflammation spreads from the vagina or cervix to the upper genital tract, with endometritis as an intermediate stage in the pathogenesis of disease.² The hallmark of the diagnosis is pelvic tenderness combined with inflammation of the lower genital tract; women with pelvic inflammatory disease often have very subtle symptoms and signs.³ Many women have clinically silent spread of infection to the upper genital tract, which results in subclinical pelvic inflammatory disease.^{1,4}

Pelvic inflammatory disease is a major concern because it can result in longterm reproductive disability, including infertility, ectopic pregnancy, and chronic pelvic pain. After the introduction of laparoscopy in the 1960s, research on pelvic inflammatory disease proliferated through the 1970s, 1980s, and 1990s, leading to major breakthroughs in the understanding of the microbial causes of the disease and its relationship to reproductive disability, as well as enabling the standardization of antimicrobial treatment. According to a national estimate, in 2001 more than 750,000 cases of pelvic inflammatory disease occurred in the United States.⁵ Over the past two decades, the rates and severity of pelvic inflammatory disease have declined in North America and western Europe.⁶⁻⁹ These declines have occurred in association with public health efforts to control Chlamydia trachomatis and Neisseria gonorrhoeae infection.^{6,10,11} Despite progress, however, pelvic inflammatory disease remains a problem because reproductive outcomes among treated patients are still suboptimal, subclinical pelvic inflammatory disease remains poorly controlled, and programs aimed at the prevention of pelvic inflammatory disease are not feasible in much of the developing world.

PATHOPHYSIOLOGY AND MICROBIAL CAUSES

Acute (\leq 30 days' duration), clinically diagnosed pelvic inflammatory disease is caused by spontaneous ascension of microbes from the cervix or vagina to the endometrium, fallopian tubes, and adjacent structures. More than 85% of infections are due to sexually transmitted cervical pathogens or bacterial vaginosis–associated microbes, and approximately 15% are due to respiratory or enteric organisms that have colonized the lower genital tract (Table 1). Subclinical pelvic inflammatory disease has causes similar to those of acute pelvic inflammatory disease and may be twice as common.^{1,12} Chronic (>30 days' duration) pelvic inflammatory disease is defined as chronic infection due to *Mycobacterium tuberculosis* or actinomyces species rather than as chronic recurrent pelvic pain, which remains common after the treatment of acute pelvic inflammatory disease.

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N Engl J Med 2015;372:2039-48. DOI: 10.1056/NEJMra1411426 Copyright © 2015 Massachusetts Medical Society.

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Table 1. Clinical Classification of Pelvic Inflammatory Disease and Likely Microbial Causes.	
Clinical Syndrome	Causes
Acute pelvic inflammatory disease (≤30 days' duration)	 Cervical pathogens (<i>Neisseria gonorrhoeae</i>, <i>Chlamydia trachomatis</i>, and <i>Mycoplasma genitalium</i>) Bacterial vaginosis pathogens (peptostreptococcus species, bacteroides species, atopobium species, leptotrichia species, <i>M. hominis</i>, <i>Ureaplasma urealyticum</i>, and clostridia species) Respiratory pathogens (<i>Haemophilus influenzae</i>, <i>Streptococcus pneumoniae</i>, group A streptococci, and <i>Staphylococcus aureus</i>) Enteric pathogens (<i>Escherichia coli</i>, <i>Bacteroides fragilis</i>, group B streptococci, and campylobacter species)
Subclinical pelvic inflammatory disease	C. trachomatis and N. gonorrhoeae
Chronic pelvic inflammatory dis- ease (>30 days' duration)	Mycobacterium tuberculosis and actinomyces species

Ascending infection from the cervix is often due to sexually acquired infections with N. gonorrhoeae or C. trachomatis. Sexually transmitted Mycoplasma genitalium has been identified as a likely cause of cervicitis, endometritis, salpingitis, and infertility, but the evidence has been inconsistent.13-15 The factors determining which cervical infections ascend to the upper genital tract have not been completely elucidated, but data from prospective studies suggest that about 15% of untreated chlamydial infections progress to clinically diagnosed pelvic inflammatory disease.16-18 The risk of pelvic inflammatory disease after gonococcal infection may be even higher. Sexual intercourse and retrograde menstruation may be particularly important in the movement of organisms from the lower to the upper genital tract.¹

Anaerobic and facultative bacteria that are found in vaginal flora have been isolated alone or with N. gonorrhoeae and C. trachomatis infection in the fallopian tubes of women with acute pelvic inflammatory disease (Table 1).1,19-23 These organisms occur in greater concentrations in association with bacterial vaginosis, a polymicrobial dysbiosis characterized by a reduction in normal vaginal lactobacilli and overgrowth of a much more complex anaerobic biofilm-associated microbiome.²⁴ Bacterial vaginosis is associated with local production of enzymes that degrade cervical mucus and associated antimicrobial peptides.3,25,26 This degradation may impair the cervical barrier to ascending infection and facilitate the spread of microorganisms to the upper genital tract.²⁷

Infection results in fibrinous or suppurative inflammatory damage along the epithelial surface of the fallopian tubes and the peritoneal surface of the fallopian tubes and ovaries, which leads to scarring, adhesions, and possibly partial or total obstruction of the fallopian tubes. The adaptive immune response plays a role in the pathogenesis of pelvic inflammatory disease because reinfection substantially increases the risk of tubalfactor infertility (i.e., the inability to conceive because of structural or functional damage to the fallopian tubes). Infection-induced selective loss of ciliated epithelial cells along the fallopian tube epithelium can cause impaired ovum transport, resulting in tubal-factor infertility or ectopic pregnancy (Fig. 1).²⁸ Peritoneal adhesions along the fallopian tubes may prevent pregnancy, and adhesions within the pelvis are related to pelvic pain.

CLINICAL MANIFESTATIONS AND DIAGNOSIS

Pelvic inflammatory disease is particularly common among sexually active young and adolescent women, who are most often treated in ambulatory clinics, physician offices, or emergency departments.^{9,29-31} The abrupt onset of severe lower abdominal pain during or shortly after menses has been the classic symptom used to identify acute pelvic inflammatory disease, although it is now well recognized that both the onset and severity of symptoms can be more ill-defined and subtle. Atypical, milder clinical manifestations have become more common as rates of N. gonorrhoeae infection have fallen.^{32,33} The symptoms associated with acute pelvic inflammatory disease include pelvic or lower abdominal pain of varying severity, abnormal vaginal discharge, intermenstrual or postcoital bleeding, dyspareunia, and

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dysuria.³⁴ Fever can occur, but systemic manifestations are not a prominent feature of pelvic inflammatory disease. Occasionally, right-upperquadrant pain suggestive of inflammation and adhesion formation in the liver capsule (perihepatitis or the Fitz-Hugh–Curtis syndrome) can accompany pelvic inflammatory disease.

A large body of evidence suggests that infection and inflammation in the upper genital tract can occur and lead to long-term reproductive complications in the absence of symptoms, a condition often called subclinical pelvic inflammatory disease.^{1,4,12} Asymptomatic infections of the upper genital tract have been well documented,35 and most women with tubal-factor infertility do not have a history of clinically diagnosed pelvic inflammatory disease, as has been observed in studies showing strong associations between infertility and serologic evidence of previous C. trachomatis or N. gonorrhoeae infection.^{36,37} Among women with tubal-factor infertility, biopsy specimens show similar pathologic tubal damage in women who have a history of pelvic inflammatory disease and those who do not.28 However, of note, in one study involving infertile women without a history of diagnosed pelvic inflammatory disease, 60% of the women with tubal-factor infertility, as compared with only 19% of those without tubal-factor infertility, reported health care visits for abdominal pain38; this suggests that many cases of pelvic inflammatory disease are missed and that clinicians should have a low threshold for considering the diagnosis.

The clinical diagnosis of pelvic inflammatory disease is based on the finding of pelvic organ tenderness, as indicated by cervical motion tenderness, adnexal tenderness, or uterine compression tenderness on bimanual examination, in conjunction with signs of lower genital tract inflammation. Signs of lower genital tract inflammation include cervical mucopus, which is visible as an exudate from the endocervix or as yellow or green mucus on a cotton-tipped swab placed gently into the cervical os (positive "swab test"); cervical friability (easily induced columnar epithelial bleeding); or increased numbers of white cells observed on saline microscopic examination of vaginal secretions (wet mount) (Fig. 2).^{39,40} Pelvic tenderness of any kind has high sensitivity (>95%) for pelvic inflammatory disease, but it has poor specificity. Findings of lower genital tract inflammation increase the specificity of the diagnosis.⁴¹ Figure S1 in the Supplementary Appendix, available with the full text of this article at NEJM.org, shows a simplified algorithm for guiding the clinical diagnosis of pelvic inflammatory disease.

Unfortunately, the clinical diagnosis of pelvic inflammatory disease is imprecise. Only about

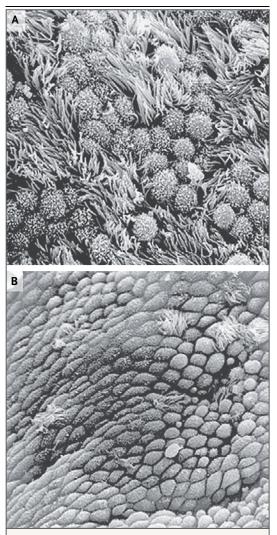


Figure 1. Pathologic Changes in the Epithelial Surface of the Fallopian Tube after Pelvic Inflammatory Disease.

Scanning electron micrographs show normal human fallopian tube epithelia (Panel A) and the epithelial surface after pelvic inflammatory disease (Panel B). Pelvic inflammatory disease causes a selective loss of ciliated epithelial cells, which interferes with intratubal ovum transport, resulting in infertility or ectopic pregnancy. Images courtesy of Dorothy L. Patton, University of Washington, Seattle.

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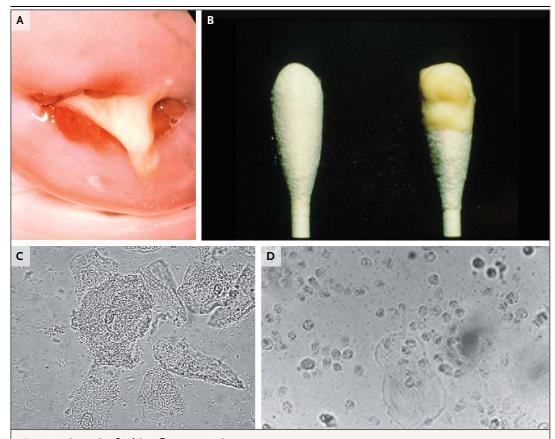


Figure 2. Diagnosis of Pelvic Inflammatory Disease.

The clinical diagnosis of pelvic inflammatory disease is based on the findings of pelvic tenderness on bimanual vaginal examination and of lower genital tract inflammation on speculum examination. Panel A shows mucopurulent endocervical discharge as seen on speculum examination. An area of endocervical columnar epithelium (ectopy) is seen on the face of the cervix. The epithelium is edematous and erythematous and bleeds easily when touched (friability). Panel B shows mucopurulent endocervical discharge as a yellow–green exudate on the tip of a Dacron swab (a positive swab test).³⁸ Panels C and D show high-power microscopic examination of vaginal fluid, with clue cells typical of bacterial vaginosis (Panel C) and increased numbers of white cells (\geq 1 per vaginal epithelial cell) (Panel D).

75% of women who have received a clinical diagnosis of pelvic inflammatory disease that is based on symptoms of pelvic tenderness and inflammation of the lower genital tract have laparoscopic confirmation of salpingitis (visualization of tubal and uterine inflammation, exudate, adhesions, or abscess).42 Although laparoscopy has been considered the standard for the diagnosis of pelvic inflammatory disease, it has high interobserver variability43 and might not detect endometritis or early tubal inflammation.44 In addition, it is an invasive surgical procedure that is not readily available in many settings and is not routinely performed, especially in women with mild-to-moderate symptoms. Transcervical endometrial aspiration with histopathological

findings of increased numbers of plasma cells and neutrophils is more commonly used to confirm the diagnosis of pelvic inflammatory disease, and these findings are often seen in association with laparoscopically confirmed salpingitis.² However, endometrial biopsy is somewhat invasive, requires skill for the pathological interpretation of the sample, and results in a delayed diagnosis.45 Transvaginal ultrasonography and magnetic resonance imaging (MRI) revealing thickened, fluidfilled tubes are available during the diagnostic workup and are highly specific for salpingitis.46,47 However, the sensitivity of ultrasonography is only fair, and although MRI has high sensitivity, it is expensive and not typically available in resource-poor settings. Power Doppler studies show-

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Table 2. First-Line Antimicrobial Treatment Recommended by the Centers for Disease Control and Prevention (CDC) for Pelvic Inflammatory Disease.*	
Out	patient regimen for mild-to-moderate pelvic inflammatory disease
Dox	ycycline (100 mg orally twice daily for 2 wk) with or without metronidazole (500 mg orally twice daily for 2 wk), plus one of the following:
(Ceftriaxone (250 mg intramuscularly in a single dose)
(Cefoxitin (2 g intramuscularly) with probenicid (1 g orally) concurrently in a single dose
(Other parenteral third-generation cephalosporin (cefotaxime or ceftizoxime)
Inpa	tient regimen for moderate-to-severe pelvic inflammatory disease with or without tubo-ovarian abscess \dag
One	of the following:
(Cefotetan (2 g intravenously every 12 hr) plus doxycycline (100 mg orally or intravenously every 12 hr)
(Cefoxitin (2 g intravenously every 6 hr) plus doxycycline (100 mg orally or intravenously every 12 hr)
(Clindamycin (900 mg intravenously every 8 hr) plus gentamicin (3 to 5 mg per kilogram of body weight intrave- nously once daily)

* Complete treatment information, including alternative regimens and additional considerations, is available at the CDC website.³³

† Transition to oral therapy can usually be initiated within 24 to 48 hours after clinical improvement, and oral therapy should be continued to complete 2 weeks of therapy.

ing increased fallopian-tube blood flow are highly suggestive of infection.^{46,48} Imaging studies may also be useful in making an alternative diagnosis, such as ovarian cyst, endometriosis, ectopic pregnancy, or acute appendicitis; these conditions can be found in 10 to 25% of women who are thought to have acute pelvic inflammatory disease.

All patients with suspected pelvic inflammatory disease should undergo cervical or vaginal nucleic acid amplification tests for N. gonorrhoeae and C. trachomatis infection; if the results are positive, the probability that pelvic inflammatory disease is present increases substantially.41 Molecular tests for M. genitalium are not yet commercially available. Vaginal fluid should be evaluated for increased numbers of white cells (more than one neutrophil per epithelial cell) and signs of bacterial vaginosis, including vaginal epithelial cells that have their cell margins obscured by attached bacteria (i.e., clue cells), an elevated pH, and an amine odor on addition of potassium hydroxide (positive "whiff" test).40 Normally, bacterial vaginosis is a noninflammatory condition, and if white cells accompany clue cells, this suggests pelvic inflammatory disease. A pregnancy test should be routinely requested to help rule out ectopic pregnancy. Serologic testing for human immunodeficiency virus (HIV) should be performed; HIV increases the risk of a tuboovarian abscess.49 An elevated erythrocyte sedimentation rate or C-reactive protein level can increase the specificity of a pelvic inflammatory disease diagnosis.⁴¹

TREATMENT

Guidelines for the treatment of pelvic inflammatory disease have been developed by the Centers for Disease Control and Prevention (CDC) on the basis of the results of clinical trials and the consensus recommendations of expert clinicians (Table 2).³³ The treatment of pelvic inflammatory disease is empirical and involves the use of broadspectrum combination regimens of antimicrobial agents to cover likely pathogens. Treatment should cover the principal pathogens, N. gonorrhoeae and C. trachomatis, regardless of the results of testing. The need to cover anaerobes has not been definitely established in randomized clinical trials, but because bacterial vaginosis is commonly found in women with pelvic inflammatory disease and anaerobes are often recovered from upper genital tract samples, antimicrobials with anaerobic coverage are recommended. Reliable coverage of *M. genitalium* is problematic, because the majority of strains are resistant to doxycycline. Moxifloxacin reliably eradicates M. genitalium¹³; however, N. gonorrhoeae has acquired quinolone resistance, and quinolone monotherapy for pelvic inflammatory disease is no longer routinely recommended.50 Substitution of azithromycin for doxycycline

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covers *M. genitalium* and simplifies dosing. However, in a recent trial of treatment for nongonococcal urethritis,⁵¹ azithromycin was found to be less reliable than doxycycline for the eradication of *C. trachomatis*, so it remains an alternative regimen.

The Pelvic Inflammatory Disease Evaluation and Clinical Health (PEACH) study showed that among women with mild-to-moderate pelvic inflammatory disease, the efficacy of cefoxitin– doxycycline therapy, with respect to both shortterm and long-term complications, was similar in inpatient and outpatient settings.⁵² The same held true for adolescents. The reasons for hospitalization for pelvic inflammatory disease currently include pregnancy, an inability to rule out competing diagnoses, severe illness combined with an inability to take oral medications, or tubal abscess.

Most patients are successfully treated as outpatients with single-dose intramuscular ceftriaxone, cefoxitin plus probenicid, or another third-generation cephalosporin (cefotaxime or ceftizoxime), followed by oral doxycycline with or without metronidazole for 2 weeks (Table 2). For hospitalized patients, therapy with cefotetan or cefoxitin (administered parenterally until 24 to 48 hours after clinical improvement) together with doxycycline and followed by doxycycline with or without metronidazole to complete 2 weeks of treatment is recommended. A regimen of clindamycin and an aminoglycoside may be particularly appropriate for patients with a tubo-ovarian abscess. Adjunctive nonsteroidal anti-inflammatory drugs do not improve the clinical outcome.53 Removal of an intrauterine device (IUD) does not hasten clinical resolution (and may delay it), and in most cases the IUD is left in place.54

LONG-TERM REPRODUCTIVE OUTCOMES

Although more than 90% of patients with pelvic inflammatory disease will have a clinical response to CDC-recommended treatment, the long-term outcome of treatment is still suboptimal. In classic studies conducted between 1960 and 1984, Westrom and colleagues followed 2501 Swedish women for several years after the women underwent laparoscopy and treatment for clinically suspected pelvic inflammatory disease; 1844 of the women (74%) had confirmed salpingitis.⁴² Infertility (i.e., an inability to conceive after 1 year of attempting to become pregnant) developed, overall, in 16% of the women with laparoscopically confirmed salpingitis, as compared with 2.7% of the women with clinically suspected pelvic inflammatory disease but no salpingitis. In addition, 9% of women with salpingitis had a subsequent ectopic pregnancy. The PEACH study provides more modern-day estimates of the risk of reproductive sequelae among 831 urban American women treated with cefoxitin and doxycycline for mild-to-moderate, clinically diagnosed pelvic inflammatory disease between 1996 and 1999.52 After 3 years of follow-up, approximately 18% of the women reported infertility, 0.6% had an ectopic pregnancy, and 29% had chronic pelvic pain (pain reported at two or more consecutive visits 3 to 4 months apart during a period of 2 to 5 years); 15% of the women had recurrent pelvic inflammatory disease.55 Both of these studies indicate that repeated episodes of pelvic inflammatory disease markedly worsen the reproductive outcomes. Of note, delayed care for pelvic inflammatory disease has also been strongly associated with worse long-term outcomes.56 It remains unclear why the long-term outcome of treated pelvic inflammatory disease remains so dismal, given the high rates of clinical response. Perhaps infection-induced damage to the fallopian tubes has occurred by the time treatment is first given. This observation, together with the frequent occurrence of subclinical pelvic inflammatory disease, have highlighted the importance of recognizing prevention of pelvic inflammatory disease as a major public heath priority.

PREVENTION

The most important public health measure for the prevention of pelvic inflammatory disease is the prevention and control of sexually transmitted infections with *C. trachomatis* or *N. gonorrhoeae*. Many high-income countries have implemented programs to screen and treat women for asymptomatic *C. trachomatis* infection, on the basis of evidence from randomized controlled trials indicating that screening for and treating cervical *C. trachomatis* infection can reduce a woman's risk of pelvic inflammatory disease by approximately 30 to 50% over 1 year.^{17,57,58} The U.S. Preventive Services Task Force, CDC, and other professional organizations recommend annual *C. trachomatis*

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screening for all sexually active women younger than 25 years of age and older women at increased risk for infection (e.g., women with multiple or new sex partners).^{33,59} These groups also recommend testing for *N. gonorrhoeae* among women at increased risk for infection (e.g., women with multiple sex partners or previous gonorrhea infection and women living in communities with a high prevalence of disease).

Comprehensive sex education, promotion of the use of condoms, and provision of condoms are cornerstones of the prevention of sexually transmitted infection globally and also have benefits for the prevention of pelvic inflammatory disease. Data from the PEACH study showed that persistent condom use during follow-up was associated with reduced risks of recurrent pelvic inflammatory disease, chronic pelvic pain, and infertility.⁶⁰ In women with pelvic inflammatory disease due to N. gonorrhoeae or C. trachomatis, reinfection and repeat pelvic inflammatory disease are common. Thus, prompt evaluation and empirical treatment of male sex partners of women with pelvic inflammatory disease or cervical infection are essential. If sex partners cannot be linked to care, expedited treatment of the partner (e.g., providing prescriptions or medications to a patient to take to her partner, without the clinician examining the partner) is a useful approach and has been shown to reduce the risk of reinfection.61

UNANSWERED QUESTIONS AND UNADDRESSED NEEDS

The National Institutes of Health recently convened a workshop to identify research needs for the improvement of the diagnosis, treatment, and prevention of pelvic inflammatory disease (Table 3).62 One of the most important needs for research regarding pelvic inflammatory disease and clinical care of women with the disease is the development of an accurate noninvasive or minimally invasive test to confirm infection of the fallopian tubes or inflammatory changes that predict long-term reproductive tract disease. Biomarkers of the immune response to C. trachomatis can predict tubal-factor infertility due to subclinical pelvic inflammatory disease.36,37 However, additional biomarkers are needed. Levels of CA-125 and E-cadherin in serum correlate with the diagnosis of acute pelvic inflammatory dis-

ease and can be used to track the response to therapy.^{63,64} Further study is needed before these assays are adopted into clinical practice. Immunohistochemical analysis and flow cytometry are being used to define specific cellular infiltrate patterns from endometrial biopsy specimens that correlate with infection.⁶⁵ Several studies that have assessed diagnostic imaging have shown the potential of MRI, transvaginal ultrasonography, and power Doppler imaging to improve the diagnosis of pelvic inflammatory disease,⁴⁷ but larger follow-up studies are needed to better define the role of these techniques in the treatment of symptomatic women and asymptomatic women with lower genital tract infection.

In recent studies in high-income populations, less than half the women with pelvic inflammatory disease have had evidence of C. trachomatis or N. gonorrhoeae infection, and the exact microbiologic cause of inflammation remains unclear.66 M. genitalium and bacterial vaginosis-associated microbes have been implicated as potential causes. Confirmatory studies are necessary to define the independent role of M. genitalium in causing pelvic inflammatory disease and longterm sequelae.13 The results of an ongoing clinical trial (ClinicalTrials.gov number, NCT01160640) evaluating the addition of metronidazole therapy to pelvic inflammatory disease regimens are expected in 2015 and should help clarify the role that organisms that cause bacterial vaginosis play in the pathogenesis of pelvic inflammatory disease. Anaerobic culture and deep sequencing methods are being used to identify specific bacterial vaginosis-associated organisms that may be more likely to cause pelvic inflammatory disease.

For financial and logistic reasons, pelvic inflammatory disease prevention programs that are based on screening are simply unavailable in most low-income and middle-income countries, where the burden of pelvic inflammatory disease may be greatest. The global epidemiologic profile of pelvic inflammatory disease has not been well defined. However, because an estimated 95.5 million C. trachomatis and N. gonorrhoeae infections occur globally among women each year⁶⁷ and approximately 15% of untreated infections lead to pelvic inflammatory disease,¹⁸ the global burden of pelvic inflammatory disease is probably substantial. The proportion of infertility that is tubal-factor infertility - and thus caused primarily by scarring from genital infec-

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Table 3. Research Needs Identified by Clinicians, Public Health Professionals, and Researchers at a 2011 National Institutes of Health Workshop.*

Characterize pathophysiological aspects of disease

Determine whether M. genitalium and bacterial vaginosis-associated organisms play a causal role in pelvic inflammatory disease and its sequelae

Determine whether histopathological endometritis correlates with subclinical pelvic inflammatory disease and its sequelae

Identify biomarkers

Identify immune and other biomarkers that correlate with pelvic inflammatory disease and its sequelae, as well as noninvasive tests to detect and measure them

Improve disease detection

Develop polymicrobial tests for lower genital tract infection Evaluate patient-administered diagnostic tests to improve case finding Determine the individual and combined predictive values of C. trachomatis and N. gonorrhoeae detection, endometrial biopsy, and imaging (MRI and ultrasonography) for the detection of pelvic inflammatory disease Determine most effective treatment Determine benefits of antimicrobial coverage for anaerobes and mycoplasma species Improve oral outpatient regimens Determine benefits of immune-modulating agents Prevent reinfection Improve mechanisms of partner treatment

* The table is adapted from Darville.62

tion — varies widely by setting. In the United States, tubal-factor infertility affects 14% of couples seeking assisted reproductive technology for infertility68; in sub-Saharan Africa, tubalfactor infertility may be present in 65 to 85% of women who seek infertility care.69,70

Most clinicians in low-income and middleincome settings rely on syndromic management (i.e., the use of genital-symptom algorithms to guide treatment) without diagnostic tests. Because most C. trachomatis and N. gonorrhoeae infections in women are asymptomatic, the majority of infections are missed. In addition, syndromic diagnosis of vaginal discharge is a poor predictor of N. gonorrhoeae and C. trachomatis cervical infection. Inexpensive, point-of-care diagnostic tests for *C.* trachomatis and *N.* gonorrhoeae that are easy to use in low-resource settings are urgently needed.⁷¹ However, the costs and complexities of screening programs may still be prohibitive. In the full text of this article at NEJM.org.

addition, the specter of cephalosporin-resistant N. gonorrhoeae looms on the horizon. Thus, the World Health Organization has concluded that the development of vaccines against C. trachomatis and N. gonorrhoeae is a critical priority for the prevention of pelvic inflammatory disease and its long-term sequelae globally.72 Progress is most advanced for C. trachomatis, for which subunit, live, and inactivated vaccines have emerged from basic research for further clinical development.73 Vaccines and other strategies to prevent pelvic inflammatory disease lie at the heart of efforts to improve women's reproductive health globally.

Dr. Brunham reports holding pending patents (WO/2013/044398; US 0027793, and PCT WO/2010/085896, CAN, US, EU, AU) related to chlamydia-specific proteins that may compose a Chlamydia trachomatis vaccine. No other potential conflict of interest relevant to this article was reported.

Disclosure forms provided by the authors are available with

REFERENCES

1. Paavonen J, Westrom L, Eschenbach D. Pelvic inflammatory disease. In: Holmes KK, Sparling PF, Stamm WE, et al., eds. Sexually transmitted diseases. 4th ed. New York: McGraw-Hill, 2008. 2. Kiviat NB, Wølner-Hanssen P, Eschen-

bach DA, et al. Endometrial histopathol-

ogy in patients with culture-proved upper genital tract infection and laparoscopically diagnosed acute salpingitis. Am J Surg Pathol 1990;14:167-75.

3. Soper DE. Pelvic inflammatory disease. Obstet Gynecol 2010;116:419-28.

4. Wiesenfeld HC, Hillier SL, Meyn LA,

Amortegui AJ, Sweet RL. Subclinical pelvic inflammatory disease and infertility. Obstet Gynecol 2012;120:37-43.

5. Sutton MY, Sternberg M, Zaidi A, St Louis ME, Markowitz LE. Trends in pelvic inflammatory disease hospital discharges and ambulatory visits, United States,

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1985-2001. Sex Transm Dis 2005;32:778-84.

6. Bender N, Herrmann B, Andersen B, et al. Chlamydia infection, pelvic inflammatory disease, ectopic pregnancy and infertility: cross-national study. Sex Transm Infect 2011;87:601-8.

7. Bohm MK, Newman L, Satterwhite CL, Tao G, Weinstock HS. Pelvic inflammatory disease among privately insured women, United States, 2001-2005. Sex Transm Dis 2010;37:131-6.

8. French CE, Hughes G, Nicholson A, et al. Estimation of the rate of pelvic inflammatory disease diagnoses: trends in England, 2000-2008. Sex Transm Dis 2011;38: 158-62.

9. Rekart ML, Gilbert M, Meza R, et al. Chlamydia public health programs and the epidemiology of pelvic inflammatory disease and ectopic pregnancy. J Infect Dis 2013;207:30-8.

10. Anschuetz GL, Asbel L, Spain CV, et al. Association between enhanced screening for Chlamydia trachomatis and Neisseria gonorrhoeae and reductions in sequelae among women. J Adolesc Health 2012;51:80-5.

11. Ross JD, Hughes G. Why is the incidence of pelvic inflammatory disease falling? BMJ 2014;348:g1538.

12. Wiesenfeld H, Cates WJ. Sexually transmitted diseases and infertility. In: Holmes KK, Sparling PF, Stamm WE, et al., eds. Sexually transmitted diseases. 4th ed. New York: McGraw-Hill, 2008.

13. Manhart LE, Broad JM, Golden MR. Mycoplasma genitalium: should we treat and how? Clin Infect Dis 2011;53:Suppl 3: S129-S142.

14. Bjartling C, Osser S, Persson K. Mycoplasma genitalium in cervicitis and pelvic inflammatory disease among women at a gynecologic outpatient service. Am J Obstet Gynecol 2012;206(6):476.e1-476.e8.

15. Bjartling C, Osser S, Persson K. The association between Mycoplasma genitalium and pelvic inflammatory disease after termination of pregnancy. BJOG 2010; 117:361-4.

16. Haggerty CL, Gottlieb SL, Taylor BD, Low N, Xu F, Ness RB. Risk of sequelae after Chlamydia trachomatis genital infection in women. J Infect Dis 2010;201: Suppl 2:S134-S155.

17. Oakeshott P, Kerry S, Aghaizu A, et al. Randomised controlled trial of screening for Chlamydia trachomatis to prevent pelvic inflammatory disease: the POPI (Prevention of Pelvic Infection) trial. BMJ 2010;340:c1642.

18. Price MJ, Ades AE, De Angelis D, et al. Risk of pelvic inflammatory disease following Chlamydia trachomatis infection: analysis of prospective studies with a multistate model. Am J Epidemiol 2013; 178:484-92.

19. Hebb JK, Cohen CR, Astete SG, Bu-

kusi EA, Totten PA. Detection of novel organisms associated with salpingitis, by use of 16S rDNA polymerase chain reaction. J Infect Dis 2004;190:2109-20.

20. Eschenbach DA, Buchanan TM, Pollock HM, et al. Polymicrobial etiology of acute pelvic inflammatory disease. N Engl J Med 1975;293:166-71.

21. Wasserheit JN, Bell TA, Kiviat NB, et al. Microbial causes of proven pelvic inflammatory disease and efficacy of clindamycin and tobramycin. Ann Intern Med 1986;104:187-93.

22. Brunham RC, Binns B, Guijon F, et al. Etiology and outcome of acute pelvic inflammatory disease. J Infect Dis 1988; 158:510-7.

23. Soper DE, Brockwell NJ, Dalton HP, Johnson D. Observations concerning the microbial etiology of acute salpingitis. Am J Obstet Gynecol 1994;170:1008-14.

24. Brotman RM. Vaginal microbiome and sexually transmitted infections: an epidemiologic perspective. J Clin Invest 2011;121:4610-7.

25. McGregor JA, French JI, Jones W, et al. Bacterial vaginosis is associated with prematurity and vaginal fluid mucinase and sialidase: results of a controlled trial of topical clindamycin cream. Am J Obstet Gynecol 1994;170:1048-59.

26. Draper DL, Landers DV, Krohn MA, Hillier SL, Wiesenfeld HC, Heine RP. Levels of vaginal secretory leukocyte protease inhibitor are decreased in women with lower reproductive tract infections. Am J Obstet Gynecol 2000;183:1243-8.

27. Ness RB, Kip KE, Hillier SL, et al. A cluster analysis of bacterial vaginosis-associated microflora and pelvic inflammatory disease. Am J Epidemiol 2005;162: 585-90.

28. Patton DL, Moore DE, Spadoni LR, Soules MR, Halbert SA, Wang SP. A comparison of the fallopian tube's response to overt and silent salpingitis. Obstet Gynecol 1989;73:622-30.

29. Schappert SM, Rechtsteiner EA. Ambulatory medical care utilization estimates for 2006. National Health Statistics Reports No. 8. Hyattsville, MD: National Center for Health Statistics, 2008:1-29 (http://www.cdc.gov/nchs/data/nhsr/nhsr08.pdf).

30. Goyal M, Hersh A, Luan X, Localio R, Trent M, Zaoutis T. National trends in pelvic inflammatory disease among adolescents in the emergency department. J Adolesc Health 2013;53:249-52.

31. Centers for Disease Control and Prevention. Sexually transmitted disease surveillance 2012. Atlanta: Department of Health and Human Services, 2013 (http://www.cdc.gov/std/stats12/surv2012.pdf)
32. Sexually transmitted infections in

Europe 1990-2010. Stockholm: European Center for Disease Prevention and Control, 2013 (http://ecdc.europa.eu/en/ publications/Publications/201206 -Sexually-Transmitted-Infections -Europe-2010.pdf).

33. Centers for Disease Control and Prevention. Pelvic inflammatory disease: STD treatment guidelines. Atlanta: Department of Health and Human Services, 2010 (http://www.cdc.gov/std/treatment/2010/pid.htm).

34. Eschenbach DA, Wölner-Hanssen P, Hawes SE, Pavletic A, Paavonen J, Holmes KK. Acute pelvic inflammatory disease: associations of clinical and laboratory findings with laparoscopic findings. Obstet Gynecol 1997;89:184-92.

35. Wiesenfeld HC, Sweet RL, Ness RB, Krohn MA, Amortegui AJ, Hillier SL. Comparison of acute and subclinical pelvic inflammatory disease. Sex Transm Dis 2005;32:400-5.

36. Brunham RC, Maclean IW, Binns B, Peeling RW. Chlamydia trachomatis: its role in tubal infertility. J Infect Dis 1985; 152:1275-82.

37. Robertson JN, Ward ME, Conway D, Caul EO. Chlamydial and gonococcal antibodies in sera of infertile women with tubal obstruction. J Clin Pathol 1987;40: 377-83.

38. Wølner-Hanssen P. Silent pelvic inflammatory disease: is it overstated? Obstet Gynecol 1995;86:321-5.

39. Brunham RC, Paavonen J, Stevens CE, et al. Mucopurulent cervicitis — the ignored counterpart in women of urethritis in men. N Engl J Med 1984;311:1-6.

40. Amsel R, Totten PA, Spiegel CA, Chen KC, Eschenbach D, Holmes KK. Nonspecific vaginitis: diagnostic criteria and microbial and epidemiologic associations. Am J Med 1983;74:14-22.

41. Peipert JF, Boardman L, Hogan JW, Sung J, Mayer KH. Laboratory evaluation of acute upper genital tract infection. Obstet Gynecol 1996;87:730-6.

42. Weström L, Joesoef R, Reynolds G, Hagdu A, Thompson SE. Pelvic inflammatory disease and fertility: a cohort study of 1,844 women with laparoscopically verified disease and 657 control women with normal laparoscopic results. Sex Transm Dis 1992;19:185-92.

43. Molander P, Finne P, Sjöberg J, Sellors J, Paavonen J. Observer agreement with laparoscopic diagnosis of pelvic inflammatory disease using photographs. Obstet Gynecol 2003;101:875-80.

44. Sellors J, Mahony J, Goldsmith C, et al. The accuracy of clinical findings and laparoscopy in pelvic inflammatory disease. Am J Obstet Gynecol 1991;164:113-20.

45. Vicetti Miguel RD, Chivukula M, Krishnamurti U, et al. Limitations of the criteria used to diagnose histologic endometritis in epidemiologic pelvic inflammatory disease research. Pathol Res Pract 2011;207:680-5.

N ENGL J MED 372;21 NEJM.ORG MAY 21, 2015

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46. Molander P, Sjoberg J, Paavonen J, Cacciatore B. Transvaginal power Doppler findings in laparoscopically proven acute pelvic inflammatory disease. Ultrasound Obstet Gynecol 2001;17:233-8.

47. Tukeva TA, Aronen HJ, Karjalainen PT, Molander P, Paavonen T, Paavonen J. MR imaging in pelvic inflammatory disease: comparison with laparoscopy and US. Radiology 1999;210:209-16.

48. Sakhel K, Benson CB, Platt LD, Goldstein SR, Benacerraf BR. Begin with the basics: role of 3-dimensional sonography as a first-line imaging technique in the cost-effective evaluation of gynecologic pelvic disease. J Ultrasound Med 2013; 32:381-8.

49. Cohen CR, Sinei S, Reilly M, et al. Effect of human immunodeficiency virus type 1 infection upon acute salpingitis: a laparoscopic study. J Infect Dis 1998;178: 1352-8.

50. Kirkcaldy RD, Bolan GA, Wasserheit JN. Cephalosporin-resistant gonorrhea in North America. JAMA 2013;309:185-7.

51. Seña AC, Lensing S, Rompalo A, et al. Chlamydia trachomatis, Mycoplasma genitalium, and Trichomonas vaginalis infections in men with nongonococcal urethritis: predictors and persistence after therapy. J Infect Dis 2012;206:357-65.
52. Ness RB, Soper DE, Holley RL, et al. Effectiveness of inpatient and outpatient treatment strategies for women with pelvic inflammatory disease: results from the Pelvic Inflammatory Disease Evaluation and Clinical Health (PEACH) Randomized Trial. Am J Obstet Gynecol 2002; 186:929-37.

53. Dhasmana D, Hathorn E, McGrath R, Tariq A, Ross JD. The effectiveness of nonsteroidal anti-inflammatory agents in the treatment of pelvic inflammatory disease: a systematic review. Syst Rev 2014;3:79.

54. Tepper NK, Steenland MW, Gaffield ME, Marchbanks PA, Curtis KM. Retention of intrauterine devices in women who acquire pelvic inflammatory disease: a systematic review. Contraception 2013;87: 655-60.

55. Ness RB, Trautmann G, Richter HE, et al. Effectiveness of treatment strategies

of some women with pelvic inflammatory disease: a randomized trial. Obstet Gynecol 2005;106:573-80.

56. Hillis SD, Joesoef R, Marchbanks PA, Wasserheit JN, Cates W Jr, Westrom L. Delayed care of pelvic inflammatory disease as a risk factor for impaired fertility. Am J Obstet Gynecol 1993;168:1503-9.

57. Gottlieb SL, Xu F, Brunham RC. Screening and treating Chlamydia trachomatis genital infection to prevent pelvic inflammatory disease: interpretation of findings from randomized controlled trials. Sex Transm Dis 2013;40:97-102.

58. Scholes D, Stergachis A, Heidrich FE, Andrilla H, Holmes KK, Stamm WE. Prevention of pelvic inflammatory disease by screening for cervical chlamydial infection. N Engl J Med 1996;334:1362-6.

59. Zakher B, Cantor AG, Pappas M, Daegas M, Nelson HD. Screening for gonorrhea and chlamydia: a systematic review for the U.S. Preventive Services Task Force. Ann Intern Med 2014;161:884-93.
60. Ness RB, Randall H, Richter HE, et al. Condom use and the risk of recurrent pelvic inflammatory disease, chronic pelvic pain, or infertility following an episode of pelvic inflammatory disease. Am J Public Health 2004;94:1327-9.

61. Golden MR, Whittington WL, Handsfield HH, et al. Effect of expedited treatment of sex partners on recurrent or persistent gonorrhea or chlamydial infection. N Engl J Med 2005;352:676-85.

62. Darville T. Pelvic inflammatory disease: identifying research gaps — proceedings of a workshop sponsored by Department of Health and Human Services/ National Institutes of Health/National Institute of Allergy and Infectious Diseases, November 3-4, 2011. Sex Transm Dis 2013; 40:761-7.

63. Paavonen J, Miettinen A, Heinonen PK, et al. Serum CA 125 in acute pelvic inflammatory disease. Br J Obstet Gynaecol 1989;96:574-9.

64. Tsai HT, Lee TH, Yang SF, Lin LY, Tee YT, Wang PH. Markedly elevated soluble E-cadherin in plasma of patient with pelvic inflammatory disease. Fertil Steril 2013;99:490-5.

65. Reighard SD, Sweet RL, Vicetti Miguel C, et al. Endometrial leukocyte subpopulations associated with Chlamydia trachomotis, Neisseria gonorrhoeae, and Trichomonas vaginalis genital tract infection. Am J Obstet Gynecol 2011;205(4):324.e1-324.e7.

66. Burnett AM, Anderson CP, Zwank MD. Laboratory-confirmed gonorrhea and/or chlamydia rates in clinically diagnosed pelvic inflammatory disease and cervicitis. Am J Emerg Med 2012;30:1114-7.

67. Global incidence and prevalence of selected curable sexually transmitted infections — 2008. Geneva: World Health Organization, 2012 (http://www.who.int/ reproductivehealth/publications/rtis/ stisestimates/en/index.html).

68. Centers for Disease Control and Prevention, American Society for Reproductive Medicine, Society for Assisted Reproductive Technology. 2011 Assisted reproductive technology fertility clinic success rates report. Atlanta: Department of Health and Human Services, 2013 (http://www.cdc.gov/art/pdf/2011-report/fertility-clinic/art_2011_clinic_report-full.pdf).

69. Cates W, Farley TM, Rowe PJ. Worldwide patterns of infertility: is Africa different? Lancet 1985;2:596-8.

70. Dhont N, van de Wijgert J, Vyankandondera J, Busasa R, Gasarabwe A, Temmerman M. Results of infertility investigations and follow-up among 312 infertile women and their partners in Kigali, Rwanda. Trop Doct 2011;41:96-101.

71. Peeling RW. Applying new technologies for diagnosing sexually transmitted infections in resource-poor settings. Sex Transm Infect 2011;87:Suppl 2:ii28-ii30.

72. Broutet N, Fruth U, Deal C, Gottlieb SL, Rees H. Vaccines against sexually transmitted infections: the way forward. Vaccine 2014;32:1630-7.

73. Brunham RC, Rappuoli R. Chlamydia trachomatis control requires a vaccine. Vaccine 2013;31:1892-7.

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